

Review Article

Arterial Hypertension and Interleukins: Potential Therapeutic Target or Future Diagnostic Marker?

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Hypertension as a multifactorial pathology is one of the most important cardiovascular risk factors, affecting up to 30-40% of the general population. Complex immune responses are involved in the inflammatory mechanism of hypertension, with evidence pointing to increased inflammatory mediators even in prehypertensive patients. Increased vascular permeability, thrombogenesis, and fibrosis, effects that are associated with sustained hypertension, could be attributed to chronic inflammation. Chronic inflammation triggers endothelial dysfunction via increased production of ROS through proinflammatory cytokines. Increased serum level of proinflammatory cytokines such as IL-1 β , IL-6, IL-8, IL-17, IL-23, TGF β , and TNF α in hypertensive patients has been associated with either increased blood pressure values and/or end-organ damage. Moreover, some cytokines (i.e., IL-6) seem to determine a hypertensive response to angiotensin II, regardless of blood pressure values. Understanding hypertension as an inflammatory-based pathology gives way to new therapeutic targets. As such, conventional cardiovascular drugs (statins, calcium channels blockers, and ACEIs/ARBs) have shown additional anti-inflammatory effects that could be linked to their blood pressure lowering properties. Moreover, anti-inflammatory drugs (mycophenolate mofetil) have been shown to decrease blood pressure in hypertensive patients or prevent its development in normotensive individuals. Further research is needed to evaluate whether drugs targeting hypertensive-linked proinflammatory cytokines, such as monoclonal antibodies, could become a new therapeutic option in treating arterial hypertension.

1. Introduction

According to WHO, cardiovascular diseases (CVDs) add disability-adjusted life years and, in 2015, caused 17.7 million deaths [1]. Arterial hypertension (HTN) is a major CVDs risk factor and multifactorial disease, affecting 30-40% of the population and causing 7.5 million deaths worldwide [2]. Despite numerous (non)pharmacological measures to prevent it/slow it down, HTN prompts 62% of strokes and 38% of heart diseases in developing countries [3]. Increasing evidence reveals HTN as a chronic inflammatory state [4, 5]. Whether inflammation contributes to HTN or HTN

generates systemic inflammation remains to be seen. Inflammatory cytokine basal levels (IL-1 β , IL-6, IL-8, IL-17, IL-23, TGF β , and TNF α) are higher in hypertensive patients. These cytokines (IL-6) trigger hypertensive responses to angiotensin II infusion even in normotensive individuals, which presents new potential therapeutic targets [6–8].

2. Inflammation in Arterial Hypertension

A key component in the pathophysiology of HTN is inflammation [9–11]. Not only does it determine HTN development and/or progression, but it also leads to end-organ damage. Metabolic/chemical, mechanical (wall stretch), or infectious endothelial aggressions trigger complex immune reactions, leading to a proinflammatory state [12, 13]. Inflammation, in turn, promotes endothelial dysfunction and atherosclerosis through reactive oxygen species (ROS), a downstream product of cellular and soluble immune factors [14–16]. Consequently, ROS stimulates proinflammatory cytokine secretion, increasing IL-6 expression and decreasing NO availability [8]. Studies have shown that inhibition of these ROS led to blood pressure reduction through endothelial function improvement via increased nitric oxide (NO) production [8, 9, 16].

2.1. RAS and Proinflammatory Cytokines. The implication of renin-angiotensin-angiotensinogen system (RAS) in the pathogenesis of HTN has been long known.

Interestingly, several immune cells (T lymphocytes, dendritic cells, and macrophages) express angiotensin 1 receptors (AT1R). By binding to AT1R, angiotensin II determines immune cells differentiation and subsequent proinflammatory cytokine production, such as IL-6, IFN- γ , and TNF α [5]. In addition, by acting on P-selectins and adhesion molecules, it increases leukocytes adhesion and migration. Moreover, angiotensin II impacts the immune system even in the absence of vasoconstrictor effects. This may explain the role of RAAS in HTN pathogenesis as an inflammatory disease [17]. In fact, it seems that angiotensin II contributes not only to HTN development, but also to HTN-mediated organ damage. In turn, proinflammatory cytokines, such as TNF- α , determine increased angiotensin converting enzyme (ACE) production, which contributes to inflammatory-mediated HTN [11, 15, 16].

Another pathway through which angiotensin II contributes to inflammation is by stimulating NADH and NADPH oxidase, determining increased reactive oxygen species (ROS), including superoxide production. Moreover, IL-6, stimulated by angiotensin II, in turn determines increased NADH and NADPH production, altering vascular permeability, constriction, and fibrosis degree [18–20].

2.2. ROS Regulation via Inflammation. It is known from preclinical models that the imbalance between reactive oxygen species production and degradation is involved in the HTN mechanisms. Because excessive ROS enhances cellular processes like differentiation and apoptosis and controls vascular tone and endothelial function, it contributes to endothelial dysfunction [21]. Increased ROS generation and reduced antioxidants levels (nitric oxide) lead to oxidative stress. ROS production involves both cellular and mitochondrial levels, with the latter being the main endogenous source. ROS molecules including xanthine oxidoreductase, uncoupled NO synthase (NOS), nicotinamide adenine dinucleotide phosphate (NADPH) nitric xanthine oxidase (NOX), and mitochondrial respiratory enzymes play a role in the HTN development [21, 22]. The production of mitochondrial ROS depends on the activation of the mitochondria ATP-sensitive potassium channels (mKATP), opening of mitochondrial permeability transition pore (mPTP), and the pH gradient in

the inner membrane. Through mitochondrial ATP synthesis function, mitochondrial respiratory chain O_2 is converted by SOD2 (superoxide dismutase) to H2O2, which is a neutral molecule. Authors have shown that mitochondrial ROS enhances NADPH production and vice versa. Therefore, mitochondrial ROS generation, as a continuous feed-forward cycle, is redox-dependent.

Both end-organ dysfunction and HTN could be a result of the mitochondrial oxidative stress, which is the outcome of overproduction of mitochondrial superoxide and reduced SOD2 function. Interchanges between mitochondrial oxidases and NOXs are highly implicated in cellular ROS production [23].

Endothelial and plasma xanthine oxidase (XO) have been linked to HTN induced atherosclerosis and associated end-organ-damage [24]. Increased oxidative stress enhances RAAS activation in hypertensive subjects. This leads to prolonged redox signaling and reduced NO bioavailability in renal microvasculature [25]. Emerging evidence proposed that inflammatory cytokines such as IL-6 influence eNOS activity and expression, NADPH oxidase with subsequent impact on NO and superoxide levels [11, 22, 26]. Studies have shown that accessory proteins such as p22phox, p47phox, p67phox and Rac1 and NOX (the main catalytic of NADPH oxidase) stimulate oxidase activity and superoxide generation. Besides NADPH oxidase, other enzymatic endothelial superoxide sources are cytochrome P450, uncoupled eNOS, cyclooxygenase, and xanthine oxidase [27].

The proinflammatory state of hypertensive patients is supported by many studies that revealed increased serum levels of C-reactive protein [16, 28–31]. Moreover, these patients had histological arterial wall inflammation attributed to proinflammatory cytokines secretion [15, 16], insulin resistance, and increased atherogenic lipoprotein levels [29–31].

2.3. Endothelial Dysfunction in Hypertensive Patients. The innermost layer of the blood vessels, the endothelium, was first considered a passive barrier between blood and vascular wall. Bunting et al. demonstrated in 1976 [29] that the arterial wall can synthetize both endothelium-derived relaxing factors (EDRF) like NO, prostacyclin (PGI₂), and hydrogen sulfide (H2S) and constrictor factors including angiotensin II (Ang II) and endothelin-1 (ET-1) [30].

Cardiovascular risk factors such as dyslipidaemia, obesity, and diabetes, through their inflammatory state, promote endothelial dysfunction. Mast cells, T lymphocytes, dendritic cells, activated neutrophils, and platelets interact to produce an inflammatory response, with increased production of proinflammatory cytokines, ROS, and adhesion molecules [14, 31–33]. In some autoimmune diseases such as psoriasis or rheumatoid arthritis [34, 35] anti-endothelial antibodies cause endothelial cells to release adhesion molecule and chemokines. Elevated serum levels of IL-6 lead to increased hepatic inflammatory markers production such as CRP [17] and increase vascular permeability, cell apoptosis, and thrombosis [36] (Figure 1). Moreover, the link between endothelial dysfunction and inflammatory cytokine production has been emphasized by studies that attribute increased IL-6 production to mechanical endothelial stretch in hypertensive

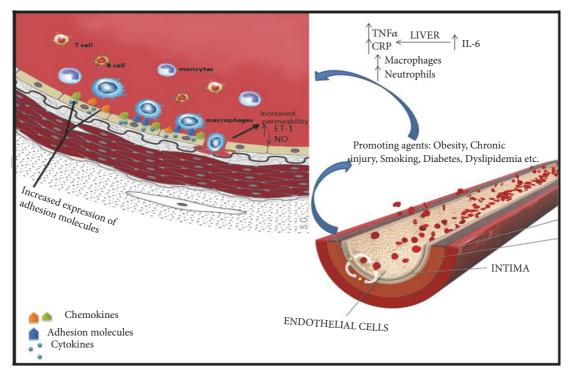


FIGURE 1: Etiology of the inflammatory process. Low level chronic inflammation increases the concentrations of markers and of inflammatory cells, leading to increased production of C-reactive protein (CRP) by the liver, in response to interleukin-6 (IL-6), which provokes a reduction in vasodilation and an increase in vascular damage. TNF-a: tumor necrosis factor alpha; IL-6: interleukin-6; CRP: C-reactive protein; NO: nitric oxide; ET-1: endothelin-1.

individuals, with a subsequent decrease in NO production [8].

Thus endothelial dysfunction is an early predictor of atherosclerosis and cardiovascular events and mortality [37], as patients with coronary artery disease [18], peripheral arterial disease [38], or heart failure [39] have shown endothelial dysfunction. Ridker et al. have pointed out to the association between increased serum levels of IL-6 and the risk of myocardial infarction [40]. Increasing evidence shows the importance of inflammation and oxidative stress in cerebral and cardiac microvascular dysfunction as well as the specific role of B-lymphocytes in cytokine production [41].

2.4. Role of Interleukins in Inflammation and HTN Development. Cytokines are involved in several cellular processes, ranging from inflammation to both tissue damage and regeneration. They induce immune cells recruitment and activation and play an important role in arthrosclerosis development [42, 43]. Proinflammatory stimuli determine mast cells cytokine production, triggering endothelial expression of adhesion molecule such as vascular cell adhesion protein 1 (VCAM-1), P- selectin, and plated activating factor (PAF) [19]. Leukocyte recruitment and adhesion promote vascular and extracellular matrix remodeling through increased fibrosis and hypertrophy with a subsequent reduction in vascular lumen diameter [44, 45].

Apart from their effect on blood pressure, interleukins, as key mediators of inflammation, seem to play a role in

end-organ damage in hypertensive individuals [19, 43, 44]. IL-6 has shown profibrotic effects, determining both vascular and myocardial fibrosis [46]. In relation to increased arterial stiffness in the context of inflammation, cardiac hypertrophy can be explained as a response to immune cell- induced mechanical changes [47, 48].

Both immune and endothelial cells contribute through their interaction to the inflammatory state in hypertensive patients. The balance between proinflammatory (IL-1 β , IL-6 IL-12, IL-18, IL-17, or IL-23) and anti-inflammatory cytokines (IL-4, IL-10) is tightly regulated and directly involved in CVDs, not only in HTN development per se, but also in mediating hypertensive end-organ damage such as ventricular remodeling and renal and cerebral involvement [20, 49]. In this inflammatory-mediated HTN process, adaptive or innate immune cells produce proinflammatory cytokines through different signaling pathways (Figure 2).

For example, IL-6R (interleukin-6 receptor) is stimulated in HTN inflammation, using STAT signaling via gp130 subunit and specific isoforms which enhance NADPH oxidase and eNOS with reduced oxide nitric levels, and increases in vascular superoxide. These actions lead to raising vascular permeability, immune cell recruitment, endothelial activation, and dysfunction. Interestingly stimulated soluble IL-6R, which reacts via membrane-anchored metalloproteinase ADAM10/ADAM17 signaling, leads to endothelial production of monocyte chemoattractant protein-1 (MCP-1). Moreover, other cytokines like IL-17 which are mainly

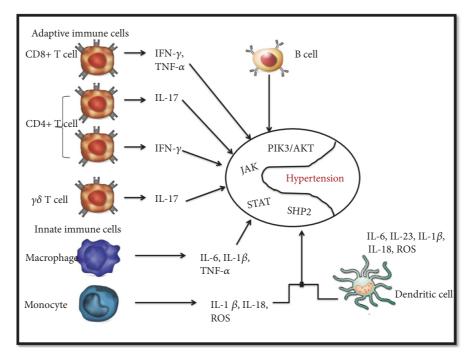


FIGURE 2: The inflammatory-mediated HTN process, immune cells via different signaling pathways. The known routes, like signal transducer and activator of transcription -1, -3, and -5 (STAT), janus-associated kinases (JAK), domain containing phosphatase (SHP2) or extracellular signal related kinase and phosphatidylinositol-3-kinase (PIK3/AKT), depend on the specific interleukin production.

produced by CD4 T cells or IFN- γ exert their inflammatory effect via similar pathways [50].

2.5. IL-1 and Hypertension. IL-1 is considered to be an "early-response" cytokine, involved in energy homeostasis and inflammation, connected to metabolism mechanisms [51]. Recent observations linked elevated levels of CRP as an indirect marker of IL-1 activity in the context of lowgrade inflammation to HTN development [52]. IL-1 pathway seems to play an important role in atherosclerosis, with IL-1 α and/or β promoting the expression of VCAM-1, ICAM-1, and E-selectin [53], with increased endothelial cell permeability, adhesion molecules expression [54]. Furthermore, endothelin-mediated vasoconstriction seems to be increased by IL-1 and TNF α [55]. IL-1 β , produced by macrophages and monocytes [56], promotes the release of IL-6 and its downstream cytokine, IL-17a, while IL-18, also a member of IL-1 family, stimulates the production of IL-2, IL-12, and IFN- γ [57, 58]. In hypertensive patients, the peripheral blood monocytes (PBMCs) are preactivated with an increased release of IL-1 β and tumor necrosis factor (TNF) [59]. In chronic hypertensive patients with/without end-organ damage, like vascular/myocardial remodeling and renal dysfunction, whether the levels of IL-1 β and IL-18 are the cause or the effect of the disease remains to be seen [60]. A study conducted by Hunag et al. [61] showed that the presence of 511T allele in the promoter region of the human IL-1 β was associated with HTN development. Moreover, several studies concluded that allele 2 of a variable number of tandem repeats (VNTR) in the intron 2 of the IL-1 receptor antagonist (IL-1 RN) gene is linked to HTN in English [62], Australian [63],

and Caucasian population [64]. However, association of IL-1 β -511C/T and IL-1 RN 86 bp VNTR polymorphisms was not relevant in the aetiology of HTN in a study conducted on 500 Pakistani Pathan subjects [65]. Also a cross-sectional study [66] conducted on 625 Japanese suggested that TT genotype of interleukin-1 β C-31T polymorphism may have a minor role in HTN development and that this association is regulated by serum β -carotene levels. However, another previous study reported no association between -1B C-31T polymorphism and HTN in Caucasians [63]. Therefore, more studies are required to identify the role of cytokine gene variants and their immunologic pathways' contribution to HTN aetiology.

Studies suggested that IL-1 receptor antagonist plays a great role in metabolic effects and that one synonymous SNP (rs315952, Ser133Ser) of the IL1RN gene was associated with development of obesity and hypertension [67]. Therefore, a therapeutic option targeting several comorbidities may be reducing the activity or expression of Il-1 β or IL-1 receptor 1 [58, 68]. Also IL-1h from the IL-1 proinflammatory superfamily has a variety of activities, including consistent effects on the atherosclerotic cell types [69, 70]. Barbieri et al. [71] showed in 537 subjects with insulin resistance syndrome that serum levels of IL-1h and IL-1ra were the only predictors of elevated diastolic blood pressure. Liu Y et al. [6] reported that endothelial cells activation leads to increased release of IL-1h and TNF α in spontaneous hypertensive rats. Furthermore, interleukin-1h infusion caused dose-dependent vasopressor response with increased blood pressure [72].

2.6. Role of IL-6 in Inflammation and Hypertension. IL-6 is a pleiotropic cytokine, with both proinflammatory and

anti-inflammatory effects [73] and multiple physiological roles. 30% of circulating IL-6 originates in adipose tissue. IL-6 promotes B cells differentiation, T cells expansion and activation, and acute-phase response regulation. Given its effects, it is now considered an important cardiovascular risk biomarker [74, 75]. Normal concentrations are relatively low (1-5 pg/ml) but are elevated in autoimmunity, infection, or cancer [25]. Signaling via gp130, the signaling subunit of the IL-6 receptor, is also needed for cell survival, growth, and function, especially Th1 and Th2 [76]. IL-6 is essential in the generation of Th17 lymphocytes via STAT3 and promotes IL-10 production, an anti-inflammatory cytokine [26, 77]. It contributes to acute-phase response by stimulating CRP hepatic synthesis, fibrinogen, and plasminogen activator inhibitor-1 (PAI-1) [78, 79], also influencing B cells proliferation [80]. Ang II, TNF- α , ET-1, IL-1 β and tissue hypoxia/ischemia stimulate IL-6 secretion with increased ROS production [81, 82]. Several studies showed that angiotensin II infusion caused both blood pressure and IL-6 serum levels increase in hypertensive patients [83, 84].

IL-6 may be involved in the pathogenesis of HTN [40, 82, 85] through its effects on vascular inflammation and stiffness and endothelial dysfunction [29, 70, 86]. Moreover, it stimulates arterial wall collagen synthesis, inhibits its degradation, and stimulates fibrinogen production [11]. Interestingly, increased IL-6 levels have been found in atherosclerotic plaques [87]. IL-6 may even show promise as a biomarker; several studies have proposed increased IL-6 and TNF- α serum levels as independent risk factors for the development of high blood pressure in apparently healthy patients [88]. A correlation between plasma levels of IL-6 and TNF- α with coronary endothelial dysfunction was found in hypertensive patients [40, 88].

Its effects on cardiac contractility reduction [89] and LV (left ventricular) remodeling- fibrosis, hypertrophy and dilatation [90, 91], support the association between IL-6 and development of end-organ damage in hypertensive patients. Janssen et al. reported in an experimental model the association of cardiac fibrosis with increased levels of IL-6 [92]. Continuous IL-6 infusion in hypertensive rats was shown to determine diastolic dysfunction irrespective of blood pressure values [93].

Consequently, IL-6 inhibition in myocardial infarction patients led to a lesser degree of left ventricular fibrosis [94]. However, Lai et al. [95] found no myocardial hypertrophy regression in IL-6 knockout (KO) mice.

The relationship between IL-6, Ang II, and aldosterone could explain the role of IL-6 in the development of HTN and provide a new therapeutic target [55, 96–98]. Sterpetti et al. [98] revealed that Ang II-mediated vasoconstriction and endothelial dysfunction may be upregulated by IL-6. Studies suggest that the renal JAK2/JAK3 pathway with its IL-6-dependent activation may play a role in Ang II mediated HTN [99]. Regarding the involvement of innate immunity in the disease pathogenesis, it seems that the loss of T cells attenuated deoxycorticosterone acetate (DOCA)-salt hypertension leads to a subsequent decrease in renal IL-6, TNF- α , ET-1, monocyte chemoattractant protein 1 (MCP-1), and ROS [100].

The role of IL-6 in the renin-angiotensin-aldosterone system (RAAS) could also explain the renal dysfunction in hypertensive patients. Ang II enhanced IL-6 levels in the kidneys with profibrotic and ET-1 gene expression induction, suggesting a possible role in chronic kidney disease (CKD) [101]. Moreover, it seems that IL-6 has a major effect in mediating the rightward shift in the renal pressure-natriuresis relationship [83] and that Ang II dependent HTN is associated with renal IL-6/ET-1 axis [101]. Conversely, IL-6 infusion in pregnant mice enhanced angiotensinogen expression in cultured renal proximal tubular cells [102], altering renal function [103, 104]. Interestingly, in haemodialysis patients, reduced levels of albumin are associated with IL-6 concentrations and increased mortality [102].

IL-6 also plays an important role in pulmonary hypertension and fibrosis [103, 105, 106]. The association between IL-6 and hypoxia is supported by increased IL-6 mRNA pulmonary levels in hypoxia-exposed mice and the partial protection against chronic hypoxia in IL-6 KO mice [106].

Prenatal IL-6 exposure has also shown to dysregulate hypothalamic-pituitary-adrenal (HPA) axis during adulthood [107]. Chronic uterine perfusion pressure reduction in (RUPP) in pregnant rats determines an increase in arterial pressure mediated by IL-6 [104, 108]. Moreover, elevated levels of both TNF α and IL-6 were identified in women with preeclampsia [109].

IL-6 gene deletion failed to affect Ang II salt induced HTN development in mice, but prevented renal dysfunction and myocardial remodeling and inflammation [110]. However, another study associated IL-6 deletion with adverse cardiac remodeling and fibrosis [111].

G to G mutations near the transcriptional start region of IL-6 (rs1800795) increase its serum concentrations and are associated with coronary artery disease (CAD) development [112, 113]. IL-6 promoter region polymorphism seems to be linked to HTN in Japanese women [114]. Single nucleotide polymorphisms (SNPs) in the -174 and -572 positions are associated with ischemic stroke [115] and CAD [116]. R Pola et al. proposed that the 174 G/C polymorphism does not affect HTN susceptibility in an elderly Italian population [117]. However, there seems to be an association between 174G/C polymorphism and left ventricular hypertrophy in dialysis hypertensive patients [118].

Several studies, including the PEGAS study [119, 120], demonstrated that the presence of 174 G/C or CRP 1846 G/A polymorphisms in specific populations increased the risk of stroke after cardiac surgery. These results suggest that ethnicity and gender might enhance the phenotypic expression of IL gene polymorphism.

IL6 _572C4G polymorphism (rs1800796) influences fibrinogen and CRP levels with no association with HTN [121]. Moreover, IL6 _572G4C polymorphism was associated with lower IL-6 levels after coronary artery bypass grafting surgery (CABG) [122]. He Ma et al. demonstrated that IL6572 C>G polymorphism correlates with HTN development [123]. A genetic variant of this polymorphism found in southern Chinese patients is rs8192284 on the IL6 receptor gene (IL6R) which appears to be associated with lower IL-6 concentrations [124]. The C alleles of rs1800795, rs1800795-, and rs1800795 were all associated with increased CAD risk [125]. Furthermore, in a Chinese population, the 634C/G polymorphism was associated with atrial fibrillation (AF) and HTN [126]. Considering these findings, further studies are required in order to fully understand the implications of IL-6 in the pathogenesis of HTN and its end-organ damage and the possible development of new antihypertensive drugs targeting inflammatory cytokines [127].

2.7. IL-17, Il-23, and IL-17/23 Axis in Hypertension Mechanisms. IL-23 is stimulated by antigen-stimulated macrophages and dendritic cells. It enhances T helper cells development (Tn17/ThIL-17) which secrete IL-17. IL-23 acts also in an autocrine/paracrine manner by promoting the release of other proinflammatory cytokines like IL-1, IL-6, and TNF- α .

The IL-17 family is formed by 6 members ranging from IL-17A to IL-17F. Despite several monoclonal antibodies targeting IL-17A, IL-17F, or the IL-17RA being used as autoimmune disease treatment (i.e., psoriasis), their effect on blood pressure has not been established [128].

There are 3 types of Th cells: Th1 cells which produce interferon-g (IFN-g), Th2 cells which produce IL-4 and IL-5; and IL-13, a third type of Th cells that secrete IL-17A, IL-17F, and IL-22 [129, 130]. The latter have a controversial role in inflammation with both atherogenic and protective roles being observed [131].

IL-17- and IL-23-producing cells have been shown to be involved in the pathogenesis of atherosclerosis [12, 131].

IL-17, by stimulating proinflammatory cytokine production, fibroblast proliferation, and profibrotic gene expression, plays a role in remodeling after post-myocardial infarction [132]. Moreover, IL-17 also interferes with tissue damage, inflammatory vascular disease [133, 134], and CAD [135]. In turn, IL-17 induces IL-1 β , IL-6, and TNF- α secretion, also associated with endothelial dysfunction and hypertension [11, 24]. To support these findings, IL-17A inhibition normalized inflammatory aortic wall infiltrate in mice [136]. IL-17 synthesis in the aortic media is also stimulated by Ang II infusion in both mice and humans [136]. Additionally, Zahra et al. [137] reported increased IL-17 levels in chronic heart failure (CHF) patients. Studies revealed that IL-17 and Ang II correlated with refractory HTN in haemodialysis patients [138, 139]. Interestingly, IL-1 is essential for Th17 development, and whether IL-1 and IL-23 act at the same molecular level remains to be seen. Moreover, IL-17/IL-23 axis deficiency in mice quickens DOCA Ang II-induced albuminuria in hypertensive kidney injury [140].

Since IL-17A is linked to Ang II induced HTN, renal dysfunction, and renal sodium transporter modulators, studies suggest that antibodies directed against IL-17A or the IL-17RA receptor subunit may be a novel adjunct hypertension therapy [141].

2.8. Other Proinflammatory Interleukins. There are more cytokines involved in the inflammatory mechanisms of HTN.

Buemia et al. showed that in patients with essential hypertension, IL-8 and ICAM-1 significantly increased the Ca2+ dependent K+ outflow in red blood cells [142]. IL-12,

mainly secreted by macrophages, is a inducer of Th-1 type cellular immune response [143].

Moreover, IL12B $^*A/^*A$ genotype was less frequent in patients with a history of stroke and IL12B $^*A/^*C$ genotype had increased risk of stroke [120].

Authors thought IL-5, with its anti-inflammatory and prothrombotic cytokine, mediated atherogenesis [144]. Kaibe et al. showed that Il-15 serum concentration were higher in hypertensive patients with severe organ damage [145].

HTN, through its effects on the vascular wall, leads to increased macrophages and mature IL-18 cells production. The subsequent natural killer and T cell maturation together with IL-2 secretion determines IL-4, IL-10, IL-13, and IFN- γ release [146].

Also, IL-18 stimulates ROS production from mechanical stress mediated NADPH oxidase and accentuates superoxideinduced HTN, suggesting a role in Ang II induced hypertension [147]. According to Jussi et al., IL-18 gene promoter region 2137G/C polymorphism modulates HTN effect on CAD and coronary atherosclerosis development [148]. IL-18 is associated with coronary stenosis and acute fatal coronary syndromes, leading to increased interferon-g (IFN-g) and matrix metalloproteinase (MMPs) secretion, which are linked to atherosclerotic plaques instability [149].

In PRME study, higher IL-18 levels seem to predict future coronary events in healthy men irrespective of blood pressure values [150]. However, more studies are needed to assess IL-8 reliability in predicting cardiovascular events.

IL-22 is another member related to IL-10 superfamily, with both pro- and anti-inflammatory properties [151]. The CC genotype of IL-22 gene polymorphism may be an independent risk factor in hypertensive patients with CAD [152], while GG genotype had a protective effect. IL-22 (rs1179251) polymorphism in the chromosome 12 (12q15, intron 4) may be associated with HTN and/or CAD [153].

A more recently discovered member of IL-1 family is IL-33. If Rana et al. suggested that IL-33/ST2 signaling may have a possible protective role in heart failure, no studies have investigated IL-33 in HTN [154].

2.9. Anti-Inflammatory Cytokines (IL-4, IL10) and Hypertension. IL-10 is known to be the main anti-inflammatory cytokine, produced by Th1, Th2, Th17, and epithelial cells and keratinocytes. It binds to its receptor, activating the IL-10/JAK1/STAT3 cascade. Hence, STAT3 phosphorylation is crucial for IL-10 pathway [155]. Its anti-inflammatory effects consist of suppressing IL-1, IL-6, IL-12 and TNF α , HLA class II, and adhesion molecules. It upregulates a tissue inhibitor of MMP-1 expression, leading to atherosclerotic plaque stabilization [156].

IL-10 could prevent aortic endothelial dysfunction caused by TNF α in mice [157]. In a another experimental transplant model, IL-10 deficiency was shown to accelerate atherosclerosis development and increase allograft refection in mice [158]. Also, IL-10 deficiency exacerbates Ang II induced vascular dysfunction [159]. Allograft survival in these models was prolonged after treatment with either rIL-10 or rIL-4 [160].

TABLE 1: Cytokines, cytokine receptors, and	their vascular impact.	HTA-arterial hypertension,	ATS-atherosclerosis, ST-stroke, IM-
myocardium infarction, CHD-coronary heart d	isease, AF-atrial fibrillat	tion, CH-cardiac hypertrophy	, LVD-left ventricule dilatation, HTP-
pulmonary hypertension, UA-unstable angina, G	CHF-chronic heart failur	re.	

Interleukine	Receptor	Cell source	Cell Target	Cardiovascular Impact
IL-1α,β	Type I IL-1r, Type II IL-1r	Monocytes/macrophage, fibroblast, endothelial cells, B cells, epithelial cells including thymic epithelium.	All cells	HTA [52, 58], ATS [53, 54, 60], IL-1β polymorphism and HTA [61–64, 66, 67], ST [73]
IL-4	IL-4 α , common γ	Mast cells, T cells, basophils.	Endothelial cells, T cells, B cells fibroblast, NK-cells, monocytes, macrophages	Anti-inflammatory action on T cells [161]
IL-6	IL-6r, gp130	fibroblast, endothelial, Mono- cytes/macrophages, most epithelial cells including thymic epithelium.	Hepatocytes, macrophages, monocytes, T cells, B cells, epithelial cells	HTA [40, 82, 85, 88], ATS [106], IM [30], CHD [112, 113, 125], AF [126], CH [90], LVD [91], HTP [103, 105, 106], ST [119, 120]
IL-10	IL-10r	T cells, B cells, monocytes macrophages, keratinocytes, mast cells	T cells, B cells, NK cells, mast cells, monocytes macrophages	Anti-inflammatory action on T cells [120, 156, 157]
IL-17	IL-17r	CD4+ T cells	Endothelium, epithelium, fibroblast, macrophages	HTA [11], ATS [12, 131], IM and UA [135], CHF [137]
IL-23	IL-12Rb1/IL23R	Macrophages, other cell types	T cells	ATS [12, 131]

IL10 gene -627 C/A polymorphism was associated with HTN in a Tatar ethnic group, with the $^{*}C/^{*}C$ genotype preventing HTN development [120].

Another pleiotropic cytokine secreted by T and B cells is IL-4. It can also be detected at the feto-maternal interface. It inhibits inflammatory cytokine production, increases MHC class II and CD23, and promotes immunoglobulin E and G1 production [160]. Piyali et al. showed that IL-4/IL-10 cotreatment during gestation in mice with preeclampsia normalized blood pressure and endothelial function and decreased the IL-6, IFN γ , TNF α , and TGF β levels [161].

Cytokines play an essential role in the inflammatory processes. As their role in the development or progression of cardiovascular diseases such as HTN or atherosclerosis (Table 1) continues to be researched, new possible therapeutic paths concerning these interleukins may be discovered.

3. Inflammation as a Potential Therapeutic Target in Arterial Hypertension

There has been consistent evidence over the past years that inflammation is a key element in the pathophysiology of hypertension, leads to, progression, and development of endorgan damage. While hypertensive patients have been shown to present with increased levels of proinflammatory cytokines such as IL-1 β , IL-6, IL-8, IL-17, and TNF α , antihypertensive treatment seems to determine a reduction in their concentration.

Considering inflammation, a new therapeutic target in hypertension is an appealing idea, especially due to its role in the pathophysiology of the disease, but more importantly, in developing end-organ damage and/through profibrotic states.

3.1. Cardiovascular Drugs Targeting Inflammation. Several drugs already used in cardiovascular pathologies have been shown to lower serum inflammatory cytokines level [162].

3.2. Statins. Statins are the most frequently used lipidlowering agents, both in primary and secondary prevention of cardiovascular events [162–165]. Apart from their effect on serum cholesterol and nonsteroidal isoprenoids synthesis through the inhibition of 3-hydroxyl-3-methyl-glutaryl coenzyme A reductase (HMGCR), they present a range of pleiotropic effects such as plaque stabilization, antithrombotic and anti-inflammatory effects, and endothelial function enhancement [165]. There has been a debate on whether statins have blood pressure lowering effects and if these effects can be attributed to their ability to reduce serum inflammatory cytokines levels and to oppose endothelial dysfunction [162]. Statins downregulate angiotensin II type 1 (AT1R) receptors [165], increase nitric oxide production through NO synthase upregulation, and inhibit ET-1 synthesis and proinflammatory cytokines production by blocking NF- κ B pathway [162, 164]. They decrease high-sensitivity CRP hepatic and plasminogen activator inhibitor-1 (PAI-1) synthesis [162].

Evelien van der Meij et al. demonstrated that statin therapy has anti-inflammatory dose-dependent effects [162]. Patients treated with either simvastatin or atorvastatin showed decreased levels of proinflammatory cytokines (MCP-1, MCP-1 ligand-ICAM-1, and IL-6) in the aortic aneurysm wall as determined by quantitative real time polymerase chain reaction (QRT-PCR). IL-6 aortic wall levels correlated with MCP-1 levels. Statin therapy influenced NF κ B dependent inflammatory mediators such as IL-6, MCP-1, TNF α , and IL-1 α and β . There was no difference in antiinflammatory effects between the two statins used, and both had no effect on inflammatory cellular component.

Statin's effect on blood pressure seems to be dependent on the severity of the disease, being more pronounced with greater values of blood pressure. Some studies report a 2-5 mm Hg reduction in systolic blood pressure after statin therapy [166].

Hypertensive patients tend to have increased interleukin 1β secretion under Ang II stimulation compared to normotensive individuals. Simvastatin therapy reduced Il- 1β levels independently of lipid-lowering effects [167].

Conversely, there are studies that found statins to possess no antihypertensive effect [164]. 4-week simvastatin therapy in normocholesterolemic normotensive rats had no effects on blood pressure levels, irrespective of dose and prior β -1 adrenergic blocker (metoprolol) infusion. However, the limitations of the study included the short period of statin administration and the use of normotensive rats, as statins' blood pressure lowering effects seem to be effective in hypertensive specimens [168].

Though their hypotensive effect is controversial when given alone, some studies [169, 170] show that statins have a synergistic effect when given with either ACEI or ARBs, suggesting that their anti-inflammatory and possible antihypertensive properties may be the result of a degree of RAAS inhibition [169].

3.3. Statins May Influence Hypertensive Ventricular Remodeling. Statins' effects in hypertension are not limited to blood pressure reduction. Ventricular remodeling in hypertensive rats seem to be influenced by statin therapy with a reverseremodeling effect. Hypertensive salt-loaded rats were randomized into 4 groups: atorvastatin, losartan, atorvastatin, and peroxisome-proliferators activated receptors (PPAR- γ) inhibitor and saline [171]. Besides reducing blood pressure values, both atorvastatin and losartan decreased serum levels of hs-CRP and of proinflammatory cytokines IL-6, IL-1 β , TNF α , and TGF- β . Both drugs prevented renal-damage development in hypertensive rats with losartan reducing NF- κ B, MCP-1, and TLR-4 levels in the kidneys. Atorvastatin had beneficial effects on ventricular hypertrophy and diastolic dysfunction. The proposed anti-inflammatory mechanism of statins included PPAR-γ pathway inhibition and G-protein signaling alteration through HMG-CoA inhibition.

It was suggested that statins alter vasopressor response to norepinephrine and could inhibit aldosterone secretion through cholesterol synthesis reduction, an effect that may depend on solubility [54]. Statins may have blood pressure lowering effects, but these effects are of limited potency when administered alone, being most likely synergic with other antihypertensive agents.

Some studies propose that the antihypertensive properties of statins failed to be noted in larger trials because of the smaller doses administered [162]. Moreover, many studies agree that the blood pressure lowering effects are dose-dependent [162, 163].

3.4. Drugs Targeting the Renin-Angiotensin-Aldosterone System. Apart from statins, first-line antihypertensives affecting the renin-angiotensin-aldosterone system (RAAS), angiotensin converting enzyme inhibitors (ACEI) and aldosterone receptor blockers (ARBs), respectively, have shown anti-inflammatory effects in addition to their blood pressure lowering effects.

Valsartan has shown anti-inflammatory effects in hypertensive patients irrespective of blood pressure reduction [54, 172]. IL-1 β basal levels in hypertensive patients randomized to either 80 mg valsartan od or other antihypertensive drugs except ARB/ACE inhibitors showed no difference between the groups. Lipopolysaccharide (LPS) stimulated IL-1 β secretion increased in all patients. Patients having received valsartan showed reduced LPS stimulated IL-1 β levels. Several studies agree that hypertensive patients showed increased IL-1 β secretion under stimulation with LPS [164, 173–176]. Treatment with losartan, amlodipine, or captopril has decreased serum IL-1 β levels irrespective of blood pressure reduction.

IL-6, IL-8, TGF- β , and TNF α levels were determined in 286 hypertensive patients and CAD patients treated with captopril, atorvastatin, losartan, aspirin, clopidogrel, metoprolol, or nitrocontin in varying doses and combinations [176]. The results showed decreased IL-6 levels and increased TGF- β in patients treated with higher doses of the aforementioned drugs except for metoprolol.

There are several proposed mechanisms for ARBs and ACEIs anti-inflammatory effects. Blockage of AT1 receptors may have anti-inflammatory effects firstly by antagonizing angiotensin II and secondly by allowing AT2 receptors activation, which may enhance NO production. Angiotensin II determines stimulation of NF- κ B transcription factors by bonding to human neutrophils, thus determining increased cytokine production. Conversely, antagonizing Ang II may reduce inflammation in both hypertensive and normotensive patients [176, 177]. Carracso-Miguel JL et al. observed that captopril reduced IL-6, IL-1 β , and TNF α serum levels in spontaneously hypertensive rats most probably due to inhibition of NF- κ B [175].

3.5. Calcium Channel Blockers. Amlodipine seems to possess the same anti-inflammatory effects as ARBs and ACEIs, decreasing serum levels of proinflammatory cytokines in hypertensive patients. Matrix metalloproteinases (MMPs)

Effects or	n inflammatory cytokines Antih	ypertensive mechanisms Proposed Reference	ces
Statins	$\downarrow IL-1\beta$ $\downarrow IL-6$ $\downarrow MCP-1$ $\downarrow ICAM-1$ $\downarrow MMP-2$ $\downarrow MMP-9$ $\downarrow hs-CRP$ $\downarrow PAI-1$ $\uparrow NO$	NF- κ B inhibition AT1R downregulation HMC CoA inhibition (G protein coupled signalling inhibition) PPAR- γ inhibition Upregulates NO synthase	[162, 164, 166, 167, 174]
ARBs/ACEIs	$\downarrow \text{IL-1}\beta$ $\downarrow \text{IL-6}$ $\uparrow \text{TGF-}\beta \text{ (losartan)}$ $\uparrow \text{ NO (AT2R)}$	NF-κB inhibition AT1R downregulation Decreased ACE synthesis	[8, 56, 164]
Calcium channel blockers	$\downarrow MMP-2$ $\downarrow MMP-9$ $\downarrow IL-1\beta$ $\downarrow IL-18$ $\downarrow CRP$ $\downarrow MCP-1$ $\downarrow ICAM-1$	Protein kinase pathway (MMP-2)	[174, 176]

TABLE 2: Anti-inflammatory effects of cardiovascular drugs.

IL: interleukin; MCP: macrophage chemotactic factor-1; ICAM-1: intercellular adhesion molecule-1; MMP: matrix metalloproteinase; NO: nitric oxide; ARBs: angiotensinogen receptor blockers; ACEIs: angiotensin converting enzyme inhibitors; NF- κ B: nuclear factor κ B; AT1R: angiotensinogen type 1 receptor; HMC CoA: hydroxy-methyl-glutaryl coenzyme A; hs-CRP: high sensitivity C reactive protein.

are endopeptidases secreted by myocardial fibroblasts and inflammatory cells that promote pressure-overload induced myocardial remodeling in hypertensive patients [177, 178]. IL-1 β , angiotensin II, and ROS stimulate MMP-2 and MMP-9 secretion, which increase the extent of myocardial fibrosis and remodeling through extracellular matrix (ECM) degradation and collagen fibers production. Tissue inhibitor of matrix metalloproteinases TIMP-1 and TIMP-2 inhibits MMP-2 and MMP-9 production. The balance between the two determines the degree of ECM degradation and thus of fibrosis. Hypertensive mice treated with either amlodipine, atorvastatin, or combination of both showed decreased levels of MMP-2 and MMP-9, which is the most potent effect in the case of the combination [176]. Lercanidipine may also have additional anti-inflammatory properties, having decreased CRP, white blood cell and neutrophil counts, superoxide release, and insulin levels in hypertensive patients [179].

Table 2 summarizes the anti-inflammatory effects of cardiovascular drugs and their potential hypotensive mechanisms.

3.6. Immunosuppressants Effects on Blood Pressure Levels. HTN has been associated with inflammation. Prehypertensive patients also present with a low-grade inflammatory state. Given that conventional antihypertensive therapies have shown anti-inflammatory effects, it is questionable whether the ability to inhibit proinflammatory cytokines such as Il-6, IL-1 β , TNF α , and IL-17 could contribute to their blood pressure lowering effects. If so, the question arises as to whether anti-inflammatory drugs could lower blood pressure in hypertensive patients, thus offering a new therapeutic option in arterial hypertension.

3.7. Mycophenolate Mofetil. Mycophenolate mofetil (MMF) is an immunosuppressant that inhibits inosine monophosphate dehydrogenase, thus limiting T and B-lymphocytes proliferation through guanosine depletion. Its indications include lupus nephritis and other autoimmune glomerular diseases, prevention of allograft rejection [180]. Systemic lupus erythematous (SLE) mice treated with MMF had lower blood pressure values as compared to vehicle-treated specimens. Moreover, in SLE mice receiving treatment with MMF, CD45R⁺ B cells depletion with subsequent autoantibody production inhibition was observed. These findings are connected to a previous study that linked autoimmunity with HTN development [181]. Hypertensive patients tend to have an increased level of AT1R antibodies, α -adrenergic receptors, and L-type voltage gated calcium channels [180-183]. Interestingly, SLE is not the only autoimmune disease in which MMF influenced blood pressure; it has proven to have similar effects in psoriasis and rheumatoid arthritis hypertensive patients [182]. Moreover, MMF tends to ameliorate renal damage in hypertensive SLE patients with a reduction in albuminuria and renal lymphocytes recruitment. Therefore, it can be assumed that MMF alleviates hypertension in SLE patients firstly through the systemic anti-inflammatory effects and secondly through the ability to prevent/ameliorate renal damage. By inhibiting glomerular immune complex deposition, MMF prevents lymphocytes infiltrates and subsequent renal inflammation that supports and even may determine hypertension development. This is in accordance with other studies [183] that confirm the blood pressure lowering effects of MMF through the reduction of renal sodium transporter.

In addition to MMF, several studies indicate the antihypertensive benefits of proinflammatory cytokines inhibition. For example, IL-17R inhibition in hypertensive mice decreases fibrosis and improves cardiac function [182]. González et al. [183] showed that IL-6 hypertensive knockout mice had preserved cardiac function as determined by left ventricle ejection fraction and a lesser degree of myocardial fibrosis and inflammation. Despite not altering angiotensin II-blood pressure response, IL-6 deletion seems to prevent cardiac remodeling and subsequent dysfunction in hypertensive mice. In addition, IL-6 knockout mice showed reduced levels of albuminuria, emphasizing the role of the proinflammatory cytokine in developing end-organ damage in hypertensive patients.

3.8. Monoclonal Antibodies. In spite of the pathophysiological link between HTN and certain proinflammatory cytokines, the antihypertensive effects of monoclonal antibodies remain controversial [184]. Studies have shown that IL-6 knockout mice present with a lower tendency to develop HTN when stimulated with angiotensin II as compared with control, thus making IL-6 a potential therapeutic target. Moreover, IL-6 inhibition in hypertensive specimens has been linked to decreased myocardial fibrosis and inflammation [183].

Monoclonal antibodies directed against IL-6 receptor such as tocilizumab are approved for the treatment of rheumatoid arthritis. Side effects include infections in the context of neutropenia and hepatotoxicity [185]. Despite the lack of studies regarding the effects of tocilizumab treatment on blood pressure, Hernández-Sánchez et al. are currently evaluating tocilizumab in pulmonary arterial hypertension patients [185].

Other available monoclonal antibodies directed against proinflammatory cytokines include sirukumab and sarilumab, also IL-6R antibodies, currently in phase 3 trials.

Taking into consideration its role in hypertension pathophysiology, IL-17 is a potential therapeutic target. Being stimulated by IL-6, IL-17 has been associated with HTN and AT1R autoantibodies production during pregnancy. IL-17 suppression through IL-17 recombinant receptor infusion seems to decrease blood pressure and AT1R autoantibodies during pregnancy [186]. Secukinumab is an anti-IL-17A antibody approved for psoriasis treatment [184]. However, one of its cited adverse effects is HTN.

So far, there is no monoclonal antibody approved for HTN treatment, especially because their blood pressure lowering effect is not well established. Moreover, their use as anti-hypertensive drugs is precluded by their side effects, including leukopenia, neutropenia, infections, and neoplasia development. There is a need for the development of new, safer monoclonal antibodies that could be further tested as potent antihypertensive drugs.

4. Discussions

Given the increasing prevalence of HTN and its effects on (cardiovascular) mortality and comorbidity, there is a constant focus on better understanding of its pathogenesis [40]. It has been known that inflammation plays an essential role in HTN [16]. Chronic inflammation enhances endothelial and tissue cells functions by promoting proinflammatory cytokine synthesis: IL-1 β , IL-6, IL-8, IL-17, IL-23, TGF β , and TNF α [28]. These cytokines contribute to elevated blood pressure with structural and myogenic modifications via elevated levels of CRP, VCAM-1, NO, EDRF, PGI2, H2S, Ang II, and ET-1 release [19, 29, 30]. IL-1 β in essential hypertension enhances VCAM-1, ICAM-1, and E-selectin expression with atherosclerotic effects [53, 54]. Different ethnic studies revealed that specific plasma levels of IL-1 β polymorphisms are associated with higher blood pressure [60–64].

IL-6 could be an important cardiovascular risk biomarker [93, 102]. Likewise, various IL-6 polymorphisms remain correlated with higher HTN risk in different populations [114–126]. IL-17 and IL-23 family were linked to atherogenic effects, endothelial dysfunction, hypertension, CHF [11, 137], and Ang II induced HTN with subsequent renal injury [140]. Over the years, cytokines such IL-8, IL-12, IL-15, IL-18, and IL-22 were associated with inflammation and HTN [142–153]. Additionally, IL-4 and IL10 exert their anti-inflammatory effect by suppressing IL-1, IL-6, IL-12 and TNF, HLA II, and adhesion molecules [156].

Taking into consideration the increase in HTN prevalence and the complex pathogenesis mechanism, targeting inflammation as a possible antihypertensive therapy is a new option. Evidence showed that several cardiovascular drugs like statins, ACEI, ARBs, and calcium channel blockers lowered blood pressure and serum inflammatory cytokines concentrations [54, 162, 164, 167, 171–179]. Even if antihypertensive effects of monoclonal antibodies remain controversial, some experimental animal studies showed that IL-6 inhibition in hypertensive specimens decreased myocardial fibrosis and reduced albuminuria, emphasizing the role of the proinflammatory cytokine in developing end-organ damage [180].

Studies showed that cardiovascular drugs and immunosuppressant molecules like MMF reduced plasma levels of different interleukins in hypertensive subjects. Even if few monoclonal antibodies were proven to exert some effects, so far no monoclonal antibody has been approved for HTN treatment. These findings emphasize the role of cytokines in the pathogenesis of hypertension and end-organ damage, pointing out to a possible therapeutic target.

5. Conclusions/Future Perspectives

Treating hypertension is not limited to blood pressure control. One of the most important goals is preventing/ counteracting end-organ damage. Taking into consideration that inflammation plays an important role in developing/maintaining both hypertension and its subsequent endorgan damage, hypertensive patients may benefit from immunosuppressive therapies as a new therapeutic option. Future prospective trials are required to assess the impact of monoclonal antibodies targeting specific interleukins on blood pressure values, end-organ damage, and systemic inflammation in hypertensive patients.

Conflicts of Interest

The authors of this review declare no conflicts of interest.

References

- [1] WHO Mediacenter, "Cardiovascular diseases (CVDs) fact sheets," 2017.
- [2] World Health Organization, "Global Health Oervatory (GHO) Data," 2015.
- [3] H. Redwood, "Hypertension, society, and public policy," European Heart Journal, Suppl 9, pp. B13–B18, 2007.
- [4] R. Kranzhöfer, J. Schmidt, C. A. H. Pfeiffer, S. Hagl, P. Libby, and W. Kübler, "Angiotensin induces inflammatory activation of human vascular smooth muscle cells," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 19, no. 7, pp. 1623–1629, 1999.
- [5] B. Schieffer, E. Schieffer, D. Hilfiker-Kleiner et al., "Expression of angiotensin II and interleukin 6 in human coronary atherosclerotic plaques: potential implications for inflammation and plaque instability," *Circulation*, vol. 101, no. 12, pp. 1372– 1378, 2000.
- [6] Y. Liu, T. Liu, R. M. McCarron et al., "Evidence for activation of endothelium and monocytes in hypertensive rats," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 270, no. 6, pp. H2125–H2131, 1996.
- [7] R. Ross, "Atherosclerosis—an inflammatory disease," *The New England Journal of Medicine*, vol. 340, no. 2, pp. 115–126, 1999.
- [8] Q.-Z. Lia, Q. Denga, J.-Q. Lia, and G.-H. Yi, "Valsartan reduces interleukin-1h secretion by peripheral blood mononuclear cells in patients with essential hypertension," *Clinica Chimica Acta*, vol. 355, pp. 131–136, 2005.
- [9] D. G. Harrison, P. J. Marvar, and J. M. Titze, "Vascular inflammatory cells in hypertension," *Frontiers in Physiology*, vol. 3, article 128, 2012.
- [10] H. Peng, X.-P. Yang, O. A. Carretero et al., "Angiotensin IIinduced dilated cardiomyopathy in Balb/c but not C57BL/6J mice," *Experimental Physiology*, vol. 96, no. 8, pp. 756–764, 2011.
- [11] L. E. Bautista, L. M. Vera, I. A. Arenas, and G. Gamarra, "Independent association between inflammatory markers (Creactive protein, interleukin-6, and TNF-alpha) and essential hypertension," *Journal of Human Hypertension*, vol. 19, no. 2, pp. 149–154, 2005.
- [12] A. Tsoupras, R. Lordan, and I. Zabetakis, "Inflammation, not cholesterol, is a cause of chronic disease," *Nutrients*, vol. 10, no. 5, p. 604, 2018.
- [13] E. Petitclerc, P. E. Pouloee, and F. Marceau, "Rapid protein synthesis and turnover is involved in interleukin-l induced relaxation of the rabbit isolated mesenteric artery. Analysis of the arachidonate cascade," *Journal of Pharmacology and Experimental Therapeutics*, vol. 268, pp. 1419–1425, 1994.
- [14] T. A. Angelovich, A. C. Hearps, and A. Jaworowski, "Inflammation-induced foam cell formation in chronic inflammatory

disease," *Immunology & Cell Biology*, vol. 93, no. 8, pp. 683–693, 2015.

- [15] A. Agita and M. Thaha, "Inflammation, immunity, and hypertension," *The Indonesian Society of Internal Medicine*, vol. 49, no. 2, 2017.
- [16] W. R. Taylor, "Hypertensive vascular disease and inflammation: mechanical and humoral mechanisms," *Current Hypertension Reports*, vol. 1, no. 1, pp. 96–101, 1999.
- [17] B. C. Teixeira, A. L. Lopes, R. C. O. Macedo et al., "Inflammatory markers, endothelial function and cardiovascular risk," *Jornal Vascular Brasileiro*, vol. 13, no. 2, pp. 108–115, 2014.
- [18] T. Heitzer, T. Schlinzig, K. Krohn, T. Meinertz, and T. Münzel, "Endothelial dysfunction, oxidative stress, and risk of cardiovascular events in patients with coronary artery disease," *Circulation*, vol. 104, no. 22, pp. 2673–2678, 2001.
- [19] M. J. Mulvany, "Small artery remodelling in hypertension," *Basic & Clinical Pharmacology & Toxicology*, vol. 110, no. 1, pp. 49–55, 2012.
- [20] C. M. Ferrario and W. B. Strawn, "Role of the renin-angiotensinaldosterone system and proinflammatory mediators in cardiovascular disease," *American Journal of Cardiology*, vol. 98, no. 1, pp. 121–128, 2006.
- [21] M. S. Finkel, C. V. Oddis, T. D. Jacob, S. C. Watkins, B. G. Hattler, and R. L. Simmons, "Negative inotropic effects of cytokines on the heart mediated by nitric oxide," *Science*, vol. 257, pp. 387– 389, 1992.
- [22] S. Didion, "Cellular and oxidative mechanisms associated with interleukin-6 signaling in the vasculature," *International Journal* of Molecular Sciences, vol. 18, no. 12, p. 2563, 2017.
- [23] S. I. Dikalov and A. E. Dikalova, "Contribution of mitochondrial oxidative stress to hypertension," *Current Opinion in Nephrology and Hypertension*, vol. 25, no. 2, pp. 73–80, 2016.
- [24] B. Legein, L. Temmerman, E. A. L. Biessen, and E. Lutgens, "Inflammation and immune system interactions in atherosclerosis," *Cellular and Molecular Life Sciences*, vol. 70, no. 20, pp. 3847–3869, 2013.
- [25] B. Mroczko, M. Groblewska, M. Gryko, B. Kedra, and Szmitkowski, "Diagnostic usefulness of serum interleukin 6 (IL-6) and C-reactive protein (CRP) in the dif-ferentiation between pancreatic cancer and chronic pancreatitis," *Journal of Clinical Laboratory Analysis*, vol. 24, pp. 256–261, 2010.
- [26] L. Zhou, I. I. Ivanov, and R. Spolski, "IL-6 programs T(H)-17 cell differentiation by promoting sequential engagement of the IL-21 and IL-23 pathways," *Nature Immunology*, vol. 8, no. 9, pp. 967–974, 2007.
- [27] D. L. Lee, J. L. Wilson, R. Duan, T. Hudson, and A. El-Marakby, "Peroxisome proliferator-activated receptor-α activation decreases mean arterial pressure, plasma interleukin-6, and COX-2 while increasing renal CYP4A expression in an acute model of DOCA-salt hypertension," *PPAR Research*, pp. 1–7, 2011.
- [28] L. Sun, Y. H. Gao, D. K. Tian, J. P. Zheng, C. Y. Zhu, and Y. Ke, "Inflammation of different tissues in spontaneously hypertensive rats," *Sheng Li Xue Bao*, vol. 58, pp. 318–323, 2006.
- [29] S. Bunting, R. Gryglewski, S. Moncada, and J. R. Vane, "Arterial walls generate from prostaglandin endoperoxides a substance (prostaglandin X) which relaxes strips of mesenteric and coeliac arteries and inhibits platelet aggregation," *Prostaglandins*, vol. 12, no. 6, pp. 897–913, 1976.
- [30] I. Bernatova, "Endothelial dysfunction in experimental models of arterial hypertension: cause or consequence?" *BioMed*

Research International, vol. 2014, Article ID 598271, 14 pages, 2014.

- [31] A. B. Tsoupras, C. Iatrou, C. Frangia, and C. A. Demopoulos, "The implication of platelet activating factor in cancer growth and metastasis: potent beneficial role of PAF-inhibitors and antioxidants," *Infectious Disorders - Drug Targets*, vol. 9, no. 4, pp. 390–399, 2009.
- [32] T. Kelesidis, V. Papakonstantinou, P. Detopoulou et al., "The role of platelet-activating factor in chronic inflammation, immune activation, and comorbidities associated with HIV infection," *AIDS Reviews*, vol. 17, no. 4, pp. 191–201, 2015.
- [33] H. B. Sager and M. Nahrendorf, "Inflammation: a trigger for acute coronary syndrome," *European Journal of Nuclear Medicine and Molecular Imaging*, vol. 60, no. 2, pp. 185–193, 2016.
- [34] V. F. Panoulas, K. M. J. Douglas, H. J. Milionis et al., "Prevalence and associations of hypertension and its control in patients with rheumatoid arthritis," *Rheumatology*, vol. 46, no. 9, pp. 1477– 1482, 2007.
- [35] A. L. Neimann, D. B. Shin, X. Wang, D. J. Margolis, A. B. Troxel, and J. M. Gelfand, "Prevalence of cardiovascular risk factors in patients with psoriasis," *Journal of the American Academy of Dermatology*, vol. 55, no. 5, pp. 829–835, 2006.
- [36] S. S. Pierangeli, R. G. Espinola, X. Liu, and E. N. Harris, "Thrombogenic effects of antiphospholipid antibodies are mediated by intercellular cell adhesion molecule-1, vascular cell adhesion molecule-1, and P-selectin," *Circulation Research*, vol. 88, no. 2, pp. 245–250, 2001.
- [37] N. Gokce, J. F. Keaney Jr., L. M. Hunter et al., "Predictive value of noninvasively determined endothelial dysfunction for longterm cardiovascular events in patients with peripheral vascular disease," *Journal of the American College of Cardiology*, vol. 41, no. 10, pp. 1769–1775, 2003.
- [38] T. J. Anderson, A. Uehata, M. D. Gerhard et al., "Close relation of endothelial function in the human coronary and peripheral circulations," *Journal of the American College of Cardiology*, vol. 26, no. 5, pp. 1235–1241, 1995.
- [39] T. Heitzer, S. Baldus, Y. Von Kodolitsch, V. Rudolph, and T. Meinertz, "Systemic endothelial dysfunction as an early predictor of adverse outcome in heart failure," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 25, no. 6, pp. 1174–1179, 2005.
- [40] P. M. Ridker, N. Rifai, M. J. Stampfer, and C. H. Hennekens, "Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men," *Circulation*, vol. 101, no. 15, pp. 1767–1772, 2000.
- [41] E. Ioannidou, "Therapeutic modulation of growth factors and cytokines in regenerative medicine," *Current Pharmaceutical Design*, vol. 12, no. 19, pp. 2397–2408, 2006.
- [42] J. Ma, H. Han, L. Ma et al., "The immunostimulatory effects of retinoblastoma cell supernatant on dendritic cells," *Protein & Cell*, vol. 5, no. 4, pp. 307–316, 2014.
- [43] E. Spinas, S. K. Kritas, A. Saggini et al., "Role of mast cells in atherosclerosis: a classical inflammatory disease," *International Journal of Immunopathology and Pharmacology*, vol. 27, no. 4, pp. 517–521, 2014.
- [44] M. V. Singh, M. W. Chapleau, S. C. Harwani, and F. M. Abboud, "The immune system and hypertension," *Immunologic Research*, vol. 59, no. 1–3, pp. 243–253, 2014.
- [45] M. Ishibashi, K.-I. Hiasa, Q. Zhao et al., "Critical role of monocyte chemoattractant protein-1 receptor CCR2 on monocytes in

hypertension-induced vascular inflammation and remodeling," *Circulation Research*, vol. 94, no. 9, pp. 1203–1210, 2004.

- [46] A. Mahmud and J. Feely, "Arterial stiffness is related to systemic inflammation in essential hypertension," *Hypertension*, vol. 46, no. 5, pp. 1118–1122, 2005.
- [47] T. J. Guzik and R. M. Touyz, "Oxidative stress, inflammation, and vascular aging in hypertension," *Hypertension*, vol. 70, no. 4, pp. 660–667, 2017.
- [48] S. Laurent and P. Boutouyrie, "The structural factor of hypertension: large and small artery alterations," *Circulation Research*, vol. 116, no. 6, pp. 1007–1021, 2015.
- [49] M. M. Manhiani, J. E. Quigley, M. J. Socha, K. Motamed, and J. D. Imig, "IL6 suppression provides renal protection independent of blood pressure in a murine model of saltsensitive hypertension," *Kidney and Blood Pressure Research*, vol. 30, no. 4, pp. 195–202, 2007.
- [50] A. E. Norlander, M. S. Madhur, and D. G. Harrison, "The immunology of hypertension," *Journal of Experimental Medicine*, vol. 215, no. 1, 2018.
- [51] J. R. Erion, M. Wosiski-Kuhn, A. Dey et al., "Obesity elicits interleukin 1-mediated deficits in hippocampal synaptic plasticity," *The Journal of Neuroscience*, vol. 34, no. 7, pp. 2618–2631, 2014.
- [52] P. Libby, P. M. Ridker, and A. Maseri, "Inflammation and atherosclerosis," *Circulation*, vol. 105, no. 9, pp. 1135–1143, 2002.
- [53] P. Welsh, G. Grassia, S. Botha, N. Sattar, and P. Maffia, "Targeting inflammation to reduce cardiovascular disease risk: a realistic clinical prospect?" *British Journal of Pharmacology*, vol. 174, no. 22, pp. 3898–3913, 2017.
- [54] D. Andrzejczak, D. Górska, and E. Czarnecka, "Influence of enalapril, quinapril and losartan on lipopolysaccharide (LPS)induced serum concentrations of TNF-α, IL-1β, IL-6 in spontaneously hypertensive rats (SHR)," *Pharmacological Reports*, vol. 59, no. 4, pp. 437–446, 2007.
- [55] L. R. White, R. Juul, K. O. Skaanes, and J. Aasly, "Cytokine enhancement of endothelin ET(B) receptor-mediated contraction in human temporal artery," *European Journal of Pharmacology*, vol. 406, no. 1, pp. 117–122, 2000.
- [56] J. M. Kahlenberg and G. R. Dubyak, "Differing caspase-1 activation states in monocyte versus macrophage models of IL-1β processing and release," *Journal of Leukocyte Biology*, vol. 76, no. 3, pp. 676–684, 2004.
- [57] C. M. Cahill and J. T. Rogers, "Interleukin (IL) 1β induction of IL-6 is mediated by a novel phosphatidylinositol 3-kinasedependent AKT/IκB kinase α pathway targeting activator protein-1," *The Journal of Biological Chemistry*, vol. 283, no. 38, pp. 25900–25912, 2008.
- [58] K. H. G. Mills, L. S. Dungan, S. A. Jones, and J. Harris, "The role of inflammasome-derived IL-1 in driving IL-17 responses," *Journal of Leukocyte Biology*, vol. 93, no. 4, pp. 489–497, 2013.
- [59] F. Nemati, N. Rahbar-Roshandel, F. Hosseini, M. Mahmoudian, and M. Shafiei, "Anti-inflammatory effects of anti-hypertensive agents: influence on interleukin-1 β secretion by peripheral blood polymorphonuclear leukocytes from patients with essential hypertension," *Clinical and Experimental Hypertension*, vol. 33, no. 2, pp. 69–76, 2011.
- [60] S. M. Krishnan, C. G. Sobey, E. Latz, A. Mansell, and G. R. Drummond, "IL-1β and IL-18: inflammatory markers or mediators of hypertension?" *British Journal of Pharmacology*, vol. 171, no. 24, pp. 5589–5602, 2014.

- [61] G. Huang, T. Niu, S. Peng et al., "Association between the interleukin-1β C(-511)T polymorphism and blood pressure in a Chinese hypertensive population," *Immunology Letters*, vol. 91, no. 2-3, pp. 159–162, 2004.
- [62] S. E. Francis, N. J. Camp, R. M. Dewberry et al., "Interleukin-1 receptor antagonist gene polymorphism and coronary artery disease," *Circulation*, vol. 99, no. 7, pp. 861–866, 1999.
- [63] R. C. Y. Lin and B. J. Morris, "Association analysis of polymorphisms at the interleukin-1 locus in essential hypertension," *American Journal of Medical Genetics*, vol. 107, no. 4, pp. 311–316, 2002.
- [64] Pakistan Medical Research Council. National Health Survey of Pakistan 1990–94. Pakistan Medical Research Council, Network Publication Service: Islamabad, Pakistan, 1998.
- [65] M. R. Khawaja, F. Taj, D. Saleheen et al., "Association study of two interleukin-1 gene loci with essential hypertension in a Pakistani Pathan population," *Journal of Human Hypertension*, vol. 22, no. 1, pp. 60–62, 2008.
- [66] A. Yanagisawa, K. Suzuki, A. Kimura, Y. Ito, N. Hamajima, and T. Inoue, "Possible protective effect of serum β-carotene levels on the association between interleukin-1B C-31T polymorphism and hypertension in a Japanese population," *Clinical Nutrition*, vol. 28, no. 2, pp. 198–202, 2009.
- [67] S. Yang, "Exonic polymorphism (rs315952, Ser133Ser) of interleukin 1 receptor antagonist (IL1RN) is related to overweigh/obese with hypertension," *Journal of Exercise Rehabilitation*, vol. 10, no. 5, pp. 332–336, 2014.
- [68] J. Qi, X.-F. Zhao, X.-J. Yu et al., "Targeting interleukin-1 beta to suppress sympathoexcitation in hypothalamic paraventricular nucleus in dahl salt-sensitive hypertensive rats," *Cardiovascular Toxicology*, vol. 16, no. 3, pp. 298–306, 2016.
- [69] D. Zohlnhofer, K. Brand, K. Schipek, G. Pogatsa-Murray, A. Schomig, and F. J. Neumann, "Adhesion of monocyte very late antigen-4 to endothelial vascular cell adhesion molecule-1 induces interleukin-1β-dependent expression of interleukin-6 in endothelial cells," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 20, no. 2, pp. 353–359, 2000.
- [70] H. Kirii, T. Niwa, Y. Yamada et al., "Lack of interleukinlh decreases the severity of atherosclerosis in ApoE-deficient mice," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 23, 656 pages, 2003.
- [71] M. Barbieri, L. Ferrucci, A. M. Corsi et al., "Is chronic inflammation a determinant of blood pressure in the elderly?" *American Journal of Hypertension*, vol. 16, no. 7, pp. 537–543, 2003.
- [72] H. Takahashi, M. Nishimura, M. Sakamoto, I. Ikegaki, T. Nakanishi, and M. Yoshimura, "Effects of interleukin-lbeta on blood pressure, sympathetic nerve activity, and pituitary endocrine functions in anesthetized rats," *American Journal of Hypertension*, vol. 5, pp. 224–229, 1992.
- [73] A. Krüttgen and S. Rose-John, "Interleukin-6 in sepsis and capillary leakage syndrome," *Journal of Interferon & Cytokine Research*, vol. 32, no. 2, pp. 60–65, 2012.
- [74] P. Stenvinkel, P. Barany, O. Heimbürger, R. Pecoits-Filho, and B. Lindholm, "Mortality, malnutrition, and atherosclerosis in ESRD: what is the role of interleukin-6?" *Kidney International*, vol. 61, pp. S103–S108, 2002.
- [75] C. A. Hunter and S. A. Jones, "IL-6 as a keystone cytokine in health and disease," *Nature Immunology*, vol. 16, no. 5, pp. 448– 457, 2015.

- [76] P. C. Heinrich, I. Behrmann, S. Haan, H. M. Hermanns, G. Müller-Newen, and F. Schaper, "Principles of interleukin (IL)-6-type cytokine signalling and its regulation," *Biochemical Journal*, vol. 374, pp. 1–20, 2003.
- [77] E. V. Acosta-Rodriguez, G. Napolitani, A. Lanzavecchia, and F. Sallusto, "Interleukins 1 β and 6 but not transforming growth factor- β are essential for the differentiation of interleukin 17-producing human T helper cells," *Nature Immunology*, vol. 8, no. 9, pp. 942–949, 2007.
- [78] A. Woods, D. J. Brull, S. E. Humphries, and H. E. Montgomery, "Genetics of inflammation and risk of coronary artery disease: the central role of interleukin-6," *European Heart Journal*, vol. 21, no. 19, pp. 1574–1583, 2000.
- [79] J. Dong, S. Fujii, D. Goto et al., "Increased expression of plasminogen activator inhibitor-1 by mediators of the acute phase response: a potential progenitor of vasculopathy in hypertensives," *Hypertension Research*, vol. 26, no. 9, pp. 723– 729, 2003.
- [80] A. K. Abbas, A. H. H. Lichtman, and S. Pillai, *Cellular and Molecular Immunology*, Elsevier Health Sciences, Philadelphia, PA, USA, 1994.
- [81] S. P. Didion, "Cellular and oxidative mechanisms associated with interleukin-6 signaling in the vasculature," *International Journal of Molecular Sciences*, vol. 18, no. 12, p. 2563, 2017.
- [82] E. A. Bermudez, N. Rifai, J. Buring, J. E. Manson, and P. M. Ridker, "Interrelationships among circulating interleukin-6, Creactive protein, and traditional cardiovascular risk factors in women," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 22, no. 10, pp. 1668–1673, 2002.
- [83] B. Chamarthi, G. H. Williams, V. Ricchiuti et al., "Inflammation and hypertension: the interplay of interleukin-6, dietary sodium, and the renin-angiotensin system in humans," *American Journal of Hypertension*, vol. 24, no. 10, pp. 1143–1148, 2011.
- [84] P. Silacci, J.-M. Dayer, A. Desgeorges, R. Peter, C. Manueddu, and P.-A. Guernet, "Interleukin (IL)-6 and its soluble receptor induce TIMP-1 expression in synoviocytes and chondrocytes, and block IL-1-induced collagenolytic activity," *The Journal of Biological Chemistry*, vol. 273, no. 22, pp. 13625–13629, 1998.
- [85] S.-Q. Mao, J.-H. Sun, T.-L. Gu, F.-B. Zhu, F.-Y. Yin, and L.-N. Zhang, "Hypomethylation of interleukin-6 (IL-6) gene increases the risk of essential hypertension: a matched casecontrol study," *Journal of Human Hypertension*, vol. 31, no. 8, pp. 530–536, 2017.
- [86] M. Gibas-Dorna, D. Nowaki, and J. Piatek, "Plasma gherilin and interleukin-6 levels correlate with body mass index and arterial blood pressure in males with essentiaial hypertension," *Journal* of Physiology and Pharmacology, vol. 66, no. 3, pp. 367–372, 2015.
- [87] S. G. Lakoski, M. Cushman, D. S. Siscovick et al., "The relationship between inflammation, obesity and risk for hypertension in the Multi-Ethnic Study of Atherosclerosis (MESA)," *Journal* of Human Hypertension, vol. 25, no. 2, pp. 73–79, 2011.
- [88] M. Naya, T. Tsukamoto, K. Morita et al., "Plasma interleukin-6 and tumor necrosis factor-α can predict coronary endothelial dysfunction in hypertensive patients," *Hypertension Research*, vol. 30, no. 6, pp. 541–548, 2007.
- [89] M. S. Finkel, C. V. Oddis, T. D. Jacob, S. C. Watkins, B. G. Hattler, and R. L. Simmons, "Negative inotropic effects of cytokines on the heart mediated by nitric oxide," *Science*, vol. 257, no. 5068, pp. 387–389, 1992.
- [90] H. Hirota, K. Yoshida, T. Kishimoto, and T. Taga, "Continuous activation of gp130, a signal-transducing receptor component

for interleukin 6-related cytokines, causes myocardial hypertrophy in mice," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 92, no. 11, pp. 4862–4866, 1995.

- [91] A. T. Yan, R. T. Yan, M. Cushman et al., "Relationship of interleukin-6 with regional and global left-ventricular function in asymptomatic individuals without clinical cardiovascular disease: insights from the multi-ethnic study of atherosclerosis," *European Heart Journal*, vol. 31, no. 7, pp. 875–882, 2010.
- [92] S. P. M. Janssen, G. Gayan-Ramirez, A. Van Den Bergh et al., "Interleukin-6 causes myocardial failure and skeletal muscle atrophy in rats," *Circulation*, vol. 111, no. 8, pp. 996–1005, 2005.
- [93] G. C. Meléndez, J. L. McLarty, S. P. Levick, Y. Du, J. S. Janicki, and G. L. Brower, "Interleukin 6 mediates myocardial fibrosis, concentric hypertrophy, and diastolic dysfunction in rats," *Hypertension*, vol. 56, no. 2, pp. 225–231, 2010.
- [94] M. Kobara, K. Noda, M. Kitamura et al., "Antibody against interleukin-6 receptor attenuates left ventricular remodelling after myocardial infarction in mice," *Cardiovascular Research*, vol. 87, no. 3, pp. 424–430, 2010.
- [95] N. C. Lai, M. H. Gao, E. Tang et al., "Pressure overloadinduced cardiac remodeling and dysfunction in the absence of interleukin 6 in mice," *Laboratory Investigation*, vol. 92, no. 11, pp. 1518–1526, 2012.
- [96] M. Sano, K. Fukuda, H. Kodama et al., "Interleukin-6 family of cytokines mediate angiotensin II-induced cardiac hypertrophy in rodent cardiomyocytes," *The Journal of Biological Chemistry*, vol. 275, no. 38, pp. 29717–29723, 2000.
- [97] A. Bürger, M. Benicke, A. Deten, and H. Zimmer, "Catecholamines stimulate interleukin-6 synthesis in rat cardiac fibroblasts," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 281, no. 1, pp. H14–H21, 2001.
- [98] A. V. Sterpetti, A. Cucina, A. R. Morena et al., "Shear stress increases the release of interleukin-1 and interleukin-6 by aortic endothelial cells," *Surgery*, vol. 114, no. 5, pp. 911–914, 1993.
- [99] W. Michael, K. L. Brands Amy, and W. Edward, "Interleukin-6 knockout prevents angiotensin II hypertension: role of renal vasoconstriction and JAK2/STAT3 activation," *Hypertension*, vol. 56, no. 5, pp. 879–884, 2010.
- [100] D. L. Lee, J. L. Wilson, R. Duan, T. Hudson, and A. El-Marakby, "Peroxisome proliferator-activated receptor- α activation decreases mean arterial pressure, plasma interleukin-6, and COX-2 while increasing renal CYP4A expression in an acute model of DOCA-salt hypertension," *PPAR Research*, vol. 2011, Article ID 502631, 7 pages, 2011.
- [101] W. Zhang, W. Wang, H. Yu et al., "Interleukin 6 underlies angiotensin II-induced hypertension and chronic renal damage," *Hypertension*, vol. 59, no. 1, pp. 136–144, 2012.
- [102] R. Satou, R. A. Gonzalez-Villalobos, K. Miyata et al., "Costimulation with angiotensin II and interleukin 6 augments angiotensinogen expression in cultured human renal proximal tubular cells," *American Journal of Physiology-Renal Physiology*, vol. 295, no. 1, pp. F283–F289, 2008.
- [103] K. W. Prins, S. L. Archer, M. Pritzker et al., "Interleukin-6 is independently associated with right ventricular function in pulmonary arterial hypertension," *The Journal of Heart and Lung Transplantation*, vol. 37, no. 3, pp. 376–384, 2018.
- [104] G. Gadonski, B. B. D. LaMarca, E. Sullivan, W. Bennett, D. Chandler, and J. P. Granger, "Hypertension produced by reductions in uterine perfusion in the pregnant rat: role of interleukin 6," *Hypertension*, vol. 48, no. 4, pp. 711–716, 2006.

- [105] L. D. Maston, D. T. Jones, W. Giermakowska et al., "Interleukin-6 trans-signaling contributes to chronic hypoxia-induced pulmonary hypertension," *Pulmonary Circulation*, vol. 8, no. 3, 2018.
- [106] L. D. Maston, D. T. Jones, W. Giermakowska et al., "Central role of T helper 17 cells in chronic hypoxia-induced pulmonary hypertension," *American Journal of Physiology-Lung Cellular* and Molecular Physiology, vol. 312, no. 5, pp. L609–L624, 2017.
- [107] A. Samuelsson, C. Alexanderson, J. Mölne, B. Haraldsson, P. Hansell, and A. Holmäng, "Prenatal exposure to interleukin-6 results in hypertension and alterations in the renin-angiotensin system of the rat," *The Journal of Physiology*, vol. 575, no. 3, pp. 855–867, 2006.
- [108] B. Giovani Gadonski, D. Babbette, and LaMarca., "Hypertension produced by reductions in uterine perfusion in the pregnant rat," *Hypertension*, vol. 48, pp. 711–716, 2006.
- [109] D. F. Benyo, A. Smarason, C. W. G. Redman, C. Sims, and K. P. Conrad, "Expression of inflammatory cytokines in placentas from women with preeclampsia," *The Journal of Clinical Endocrinology & Metabolism*, vol. 86, no. 6, pp. 2505–2512, 2001.
- [110] G. E. González, N. E. Rhaleb, M. A. D'Ambrosio et al., "Deletion of interleukin-6 prevents cardiac inflammation, fibrosis and dysfunction without affecting blood pressure in angiotensin IIhigh salt-induced hypertension," *Journal of Hypertension*, vol. 33, no. 1, pp. 144–152, 2015.
- [111] I. Banerjee, J. W. Fuseler, A. R. Intwala, and T. A. Baudino, "IL-6 loss causes ventricular dysfunction, fibrosis, reduced capillary density, and dramatically alters the cell populations of the developing and adult heart," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 296, no. 5, pp. H1694– H1704, 2009.
- [112] IL6R Genetics Consortium Emerging Risk Factors Collaboration, N. Sarwar, A. S. Butterworth et al., "Interleukin-6 receptor pathways in coronary heart disease: a collaborative meta-analysis of 82 studies," *The Lancet*, vol. 379, pp. 1205–1213, 2012.
- [113] P. Deloukas, S. Kanoni, and C. Willenborg, "Large-scale association analysis identifies new risk loci for coronary artery disease," *Nature Genetics*, vol. 45, no. 1, pp. 25–33, 2013.
- [114] T. Nakajima, N. Ota, H. Yoshida, S. Watanabe, T. Suzuki, and M. Emi, "Allelic variants in the interleukin-6 gene and essential hypertension in Japanese women," *Genes & Immunity*, vol. 1, no. 2, pp. 115–119, 1999.
- [115] Y. Tong, Z. Wang, Y. Geng et al., "The association of functional polymorphisms of IL-6 gene promoter with ischemic stroke: Analysis in two Chinese populations," *Biochemical and Biophysical Research Communications*, vol. 391, no. 1, pp. 481–485, 2010.
- [116] F. Basso, G. D. Lowe, A. Rumley, A. D. McMahon, and S. E. Humphries, "Interleukin-6 - 174G>Cpolymorphism and risk of coronary heart disease in West of Scotland coronary prevention study (WOSCOPS)," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 22, pp. 599–604, 2002.
- [117] R. Pola, A. Flex, E. Gaetani, P. Pola, and R. Bernabei, "The -174 G/C polymorphism of the interleukin-6 gene promoter and essential hypertension in an elderly Italian population," *Journal* of Human Hypertension, vol. 16, no. 9, pp. 637–640, 2002.
- [118] A. Losito, K. Kalidas, S. Santoni, and S. Jeffery, "Association of interleukin-6—174G/C promoter polymorphism with hypertension and left ventricular hypertrophy in dialysis patients," *Kidney International*, vol. 64, no. 2, pp. 616–622, 2003.

- [119] H. P. Grocott, W. D. White, R. W. Morris et al., "Genetic polymorphisms and the risk of stroke after cardiac surgery," *Stroke*, vol. 36, no. 9, pp. 1854–1858, 2005.
- [120] Y. R. Timasheva, T. R. Nasibullin, A. N. Zakirova, and O. E. Mustafina, "Association of interleukin-6, interleukin-12, and interleukin-10 gene polymorphisms with essential hypertension in Tatars from Russia," *Biochemical Genetics*, vol. 46, no. 1-2, pp. 64–74, 2008.
- [121] A. Elsaid, A. F. Abdel-Aziz, and R. Elmougy, "Association of polymorphisms G(-174)C in IL-6 gene and G(-1082)A in IL-10 gene with traditional cardiovascular risk factors in patients with coronary artery disease," *Indian Journal of Biochemistry & Biophysics*, vol. 51, pp. 282–292, 2014.
- [122] D. J. Brull, H. E. Montgomery, J. Sanders et al., "Interleukin-6 gene -174G > C and -572G > C promoter polymorphisms are strong predictors of plasma interleukin-6 levels after coronary artery bypass surgery," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 21, no. 9, pp. 1458–1463, 2001.
- [123] H. Ma, G. Sun, and W. Wang, "Association between interleukin-6 572 C>G and -174 G>C polymorphisms and hypertension," *Medicine*, vol. 95, no. 2, p. e2416, 2016.
- [124] C. Q. Jiang, T. H. Lam, B. Liu et al., "Interleukin-6 receptor gene polymorphism modulates interleukin-6 levels and the metabolic syndrome: GBCS-CVD," *Obesity*, vol. 18, no. 10, pp. 1969–1974, 2010.
- [125] H. Chen, D. Shifang, X. Liu, J. Liu, W. Yun, and W. Xiayin, "Association of interleukin-6 genetic polymorphisms and environment factors interactions with coronary artery disease in a chinese han population," *Clinical and Experimental Hypertension*, vol. 40, no. 6, pp. 514–517, 2018.
- [126] J. Li, J. Song, M.-H. Jiang et al., "Interleukin-6 promoter polymorphisms and susceptibility to atrial fibrillation in Elderly Han Chinese patients with essential hypertension," *Journal of Interferon & Cytokine Research*, vol. 32, no. 11, pp. 542–547, 2012.
- [127] I. M. Seropian, S. Toldo, B. W. van Tassell, and A. Abbate, "Antiinflammatory strategies for ventricular remodeling following St-segment elevation acute myocardial infarction," *Journal of the American College of Cardiology*, vol. 63, no. 16, pp. 1593–1603, 2014.
- [128] Y. Iwakura and H. Ishigame, "The IL-23/IL-17 axis in inflammation," *The Journal of Clinical Investigation*, vol. 116, no. 5, pp. 1218–1222, 2006.
- [129] L. E. Harrington, R. D. Hatton, P. R. Mangan et al., "Interleukin 17-producing CD4⁺ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages," *Nature Immunology*, vol. 6, no. 11, pp. 1123–1132, 2005.
- [130] M. H. Lexberg, A. Taubner, A. Förster et al., "Th memory for interleukin-17 expression is stable in vivo," *European Journal of Immunology*, vol. 38, no. 10, pp. 2654–2664, 2008.
- [131] C. Erbel, T. J. Dengler, S. Wangler et al., "Expression of IL-17A in human atherosclerotic lesions is associated with increased inflammation and plaque vulnerability," *Basic Research in Cardiology*, vol. 106, no. 1, pp. 125–134, 2011.
- [132] X. Yan, T. Shichita, Y. Katsumata et al., "Deleterious effect of the IL-23/IL-17A axis and $\gamma\delta$ T cells on left ventricular remodeling after myocardial infarction," *Journal of the American Heart Association*, vol. 1, no. 5, Article ID e004408, 2012.
- [133] M. V. Singh, M. Z. Cicha, S. Kumar et al., "Abnormal CD161 + immune cells and retinoic acid receptor-related orphan receptor γt-mediate enhanced IL-17F expression in the setting of genetic hypertension," *The Journal of Allergy and Clinical Immunology*, vol. 140, no. 3, pp. 809–821, 2017.

- [134] S. Von Vietinghoff and K. Ley, "Interleukin 17 in vascular inflammation," *Cytokine & Growth Factor Reviews*, vol. 21, no. 6, pp. 463–469, 2010.
- [135] X. Cheng, X. Yu, Y. Ding et al., "The Th17/Treg imbalance in patients with acute coronary syndrome," *Clinical Immunology*, vol. 127, pp. 89–97, 2009.
- [136] S. Meena, E. Madhur Heinrich, and A. Louise, "Interleukin 17 promotes angiotensin II-induced hypertension and vascular dysfunction," *Hypertension*, vol. 55, pp. 500–507, 2010.
- [137] Z. Rahmati, A. A. Amirzargar, S. Saadati et al., "Association of levels of interleukin 17 and T-helper 17 count with symptom severity and etiology of chronic heart failure: a case-control study," *Croatian Medical Journal*, vol. 59, no. 4, pp. 139–148, 2018.
- [138] Z. Wang, W. Shi, X. Liang, W. Wang, and J. Liang, "Association of interleukin 17 / angiotensin II with refractory hypertension risk in hemodialysis patients," *African Health Sciences*, vol. 16, no. 3, pp. 766–771, 2016.
- [139] B. H. Chung, K. W. Kim, I. O. Sun et al., "Increased interleukin-17 producing effector memory T cells in the end-stage renal disease patients," *Immunology Letters*, vol. 141, no. 2, pp. 181–189, 2012.
- [140] C. F. Krebs, S. Lange, G. Niemann et al., "Deficiency of the interleukin 17/23 axis accelerates renal injury in mice with deoxycorticosterone acetate+angiotensin II-induced hypertension," *Hypertension*, vol. 63, no. 3, pp. 565–571, 2014.
- [141] M. A. Saleh, E. Allison, and S. M. Madhur, "Inhibition of interleukin-17a, but not interleukin-17F, signaling lowers blood pressure, and reduces end-organ inflammation in angiotensin II-induced hypertension," *JACC: Basic to Translational Science*, vol. 1, pp. 606–616, 2016.
- [142] M. Buemi, D. Marino, F. Floccari et al., "Effect of interleukin 8 and ICAM-1 on calcium-dependent outflow of K + in erythrocytes from subjects with essential hypertension," *Current Medical Research and Opinion*, vol. 20, no. 1, pp. 19–24, 2004.
- [143] R. Bergholdt, P. Ghandil, J. Johannesen et al., "Genetic and functional evaluation of an interleukin-12 polymorphism (IDDM18) in families with type 1 diabetes," *Journal of Medical Genetics*, vol. 41, no. 4, p. e39, 2004.
- [144] S. L. Hoong and G. Y. H. Lip, "Interleukin-15 in hypertension: Further insights into inflammation and vascular disease," *American Journal of Hypertension*, vol. 18, no. 8, pp. 1017-1018, 2005.
- [145] M. Kaibe, M. Ohishi, N. Ito et al., "Serum interleukin-15 concentration in patients with essential hypertension," *American Journal of Hypertension*, vol. 18, no. 8, pp. 1019–1025, 2005.
- [146] J. A. Gracie, S. E. Robertson, and I. B. McInnes, "Interleukin-18," *Journal of Leukocyte Biology*, vol. 73, no. 2, pp. 213–224, 2003.
- [147] C. Elbim, C. Guichard, P. M. Dang et al., "Interleukin-18 primes the oxidative burst of neutrophils in response to formylpeptides: role of cytochrome b558 translocation and n-formyl peptide receptor endocytosis," *Clinical and Vaccine Immunology*, vol. 12, no. 3, pp. 436–446, 2005.
- [148] J. A. Hernesniemi, P. J. Karhunen, N. Oksala et al., "Interleukin 18 gene promoter polymorphism: a link between hypertension and pre-hospital sudden cardiac death: the helsinki sudden death study," *European Heart Journal*, vol. 30, no. 23, pp. 2939– 2946, 2009.
- [149] J. C. Bis, S. R. Heckbert, N. L. Smith et al., "Variation in inflammation-related genes and risk of incident nonfatal myocardial infarction or ischemic stroke," *Atherosclerosis*, vol. 198, no. 1, pp. 166–173, 2008.

- [150] S. Blankenberg, G. Luc, P. Ducimetière et al., "Interleukin-18 and the risk of coronary heart disease in european men: the prospective epidemiological study of myocardial infarction (PRIME)," *Circulation*, vol. 108, no. 20, pp. 2453–2459, 2003.
- [151] G. Schett, D. Elewaut, I. B. McInnes, J.-M. Dayer, and M. F. Neurath, "How cytokine networks fuel inflammation: toward a cytokine-based disease taxonomy," *Nature Medicine*, vol. 19, no. 7, pp. 822–824, 2013.
- [152] H. Akbari, G. Asadikaram, A. Jafari et al., "Atorvastatin, losartan and captopril may upregulate IL-22 in hypertension and coronary artery disease; the role of gene polymorphism," *Life Sciences*, vol. 207, pp. 525–531, 2018.
- [153] W. P. Arend, G. Palmer, and C. Gabay, "IL-1, IL-18, and IL-33 families of cytokines," *Immunological Reviews*, vol. 223, no. 1, pp. 20–38, 2008.
- [154] R. Ghali, R. Altara, W. E. Louch et al., "IL-33 (interleukin 33)/sST2 axis in hypertension and heart failure," *Hypertension*, vol. 72, no. 4, pp. 818–828, 2018.
- [155] J. S. G. Miguez, V. Dela Justina, A. F. M. Bressan et al., "O-Glycosylation with O-linked β-N-acetylglucosamine increases vascular contraction: possible modulatory role on Interleukin-10 signaling pathway," *Life Sciences*, vol. 209, pp. 78–84, 2018.
- [156] M. Girndt and H. Köhler, "Interleukin-10: an update on its relevance for cardiovascular risk," *Nephrol Dial Transplant*, vol. 18, pp. 1976–1979, 2003.
- [157] S. M. Zemse, C. W. Chiao, R. H. P. Hilgers, and R. C. Webb, "Interleukin-10 inhibits the in vivo and in vitro adverse effects of TNF-α on the endothelium of murine aorta," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 299, no. 4, pp. H1160–H1167, 2010.
- [158] Z. F. Yang, P. Ngai, C. K. Lau et al., "Induction of long-term liver allograft survival by delayed immunosuppression is dependent on interleukin-10," *Liver Transplantation*, vol. 13, no. 4, pp. 571– 578, 2007.
- [159] V. V. Lima, S. M. Zemse, C.-W. Chiao et al., "Interleukin-10 limits increased blood pressure and vascular RhoA/Rho-kinase signaling in angiotensin II-infused mice," *Life Sciences*, vol. 145, pp. 137–143, 2016.
- [160] M. S. Mulligan, R. L. Warner, J. E. McDuffie, S. F. Bolling, J. V. Sarma, and P. A. Ward, "Regulatory role of Th-2 cytokines, IL-10 and IL-4, in cardiac allograft rejection," *Experimental and Molecular Pathology*, vol. 69, no. 1, pp. 1–9, 2000.
- [161] P. Chatterjee, V. L. Chiasson, G. Seerangan et al., "Cotreatment with interleukin 4 and interleukin 10 modulates immune cells and prevents hypertension in pregnant mice," *American Journal* of Hypertension, vol. 28, no. 1, pp. 135–142, 2015.
- [162] E. van der Meij, G. G. Koning, P. W. Vriens et al., "A clinical evaluation of statin pleiotropy: statins selectively and dosedependently reduce vascular inflammation," *PLoS ONE*, vol. 8, no. 1, Article ID e53882, 2013.
- [163] S. M. Grundy, N. J. Stone, A. L. Bailey et al., "Cholesterol clinical practice guidelines," *American College of Cardiology*, vol. 1097, no. 18, pp. 39033–39038, 2018.
- [164] Z. Sepehri, M. Masoumi, N. Ebrahimi et al., "Atorvastatin, losartan and captopril lead to upregulation of TGF- β , and downregulation of IL-6 in coronary artery disease and hypertension," *PLoS ONE*, vol. 11, no. 12, Article ID e0168312, 2016.
- [165] M. Rizzo, G. Montalto, M. Banach et al., "The effects of statins on blood pressure: current knowledge and future perspectives," *Archives of Medical Science*, vol. 8, no. 1, pp. 1–3, 2012.

- [166] Z. Shuiping, L. Quanzhong, L. Ling et al., "Simvastatin reduces interleukin-1β secretion by peripheral blood mononuclear cells in patients with essential hypertension," *Clinica Chimica Acta*, vol. 344, no. 1-2, pp. 195–200, 2004.
- [167] J. Owczarek, M. Jasińska, I. Wejman, U. Kurczewska, and D. Orszulak-Michalak, "Effects of 4-week administration of simvastatin in different doses on heart rate and blood pressure after metoprolol injection in normocholesterolaemic and normotensive rats," *Archives of Medical Science*, vol. 8, no. 1, pp. 17– 21, 2012.
- [168] A. Drapala, M. Aleksandrowicz, T. Zera et al., "The effect of simvastatin and pravastatin on arterial blood pressure, baroreflex, vasoconstrictor, and hypertensive effects of angiotensin II in Sprague-Dawley rats," *Journal of the American Society of Hypertension*, vol. 8, no. 12, pp. 863–871, 2014.
- [169] J. M. Ledingham and R. Laverty, "Effect of simvastatin given alone and in combination with valsartan or enalapril on blood pressure and the structure of mesenteric resistance arteries and the basilar artery in the genetically hypertensive rat model," *Clinical and Experimental Pharmacology and Physiology*, vol. 32, no. 1-2, pp. 76–85, 2005.
- [170] J. Zhao, Q. Cheng, Y. Liu, G. Yang, and X. Wang, "Atorvastatin alleviates early hypertensive renal damage in spontaneously hypertensive rats," *Biomedicine & Pharmacotherapy*, vol. 109, pp. 602–609, 2019.
- [171] A. Drapala, M. Sikora, M. Ufnal et al., "Statins, the reninangiotensin-aldosterone system and hypertension - A tale of another beneficial effect of statins," *JRAAS - Journal of the Renin-Angiotensin-Aldosterone System*, vol. 15, no. 3, pp. 250–258, 2014.
- [172] D. Andrzejczak, D. Górska, and E. Czarnecka, "Influence of amlodipine and atenolol on lipopolysaccharide (LPS)-induced serum concentrations of TNF-α, IL-1β, IL-6 in spontaneously hypertensive rats (SHR)," *Pharmacological Reports*, vol. 58, no. 5, pp. 711–719, 2006.
- [173] J. Zhang, N. P. Rudemiller, M. B. Patel et al., "Interleukin-1 receptor activation potentiates salt reabsorption in angiotensin II-induced hypertension via the NKCC2 Co-transporter in the nephron," *Cell Metabolism*, vol. 23, no. 2, pp. 360–368, 2016.
- [174] J. Lu, J. Hao, H. Du et al., "Amlodipine and atorvastatin improved hypertensive cardiac remodeling through regulation of MMPs/TIMPs in SHR rats," *Cellular Physiology and Biochemistry*, vol. 39, no. 1, pp. 47–60, 2016.
- [175] D. L. Mattson, L. James, E. A. Berdan, and C. J. Meister, "Immune suppression attenuates hypertension and renal disease in the Dahl salt-sensitive rat," *Hypertension*, vol. 48, no. 1, pp. 149–156, 2006.
- [176] R. Farah, R. Khamisy-Farah, R. Shurtz-Swirski et al., "Calcium channel blocker effect on insulin resistance and inflammatory markers in essential hypertension patients," *International Angiology*, vol. 32, no. 1, pp. 85–93, 2013.
- [177] C. de Miguel, S. Das, H. Lund, and D. L. Mattson, "T lymphocytes mediate hypertension and kidney damage in Dahl salt-sensitive rats," *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, vol. 298, no. 4, pp. R1136–R1142, 2010.
- [178] E. I. Boesen, D. L. Williams, J. S. Pollock, and D. M. Pollock, "Immunosuppression with mycophenolate mofetil attenuates the development of hypertension and albuminuria in deoxycorticosterone acetate-salt hypertensive rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 37, no. 10, pp. 1016– 1022, 2010.

- [179] M. C Villarroel, M. Hidalgo, and A. Jimeno, "Mycophenolate mofetil: an update," *Drugs Today (Barc)*, vol. 45, no. 7, pp. 521– 532, 2009.
- [180] B. Rodríguez-Iturbe, Y. Quiroz, M. Nava et al., "Reduction of renal immune cell infiltration results in blood pressure control in genetically hypertensive rats," *American Journal of Physiology-Renal Physiology*, vol. 282, no. 2, pp. F191–F201, 2002.
- [181] A. D. Moes, D. Severs, K. Verdonk et al., "Mycophenolate mofetil attenuates DOCA-salt hypertension: effects on vascular tone," *Frontiers in Physiology*, vol. 9, p. 578, 2018.
- [182] E. B. Taylor and M. J. Ryan, "Immunosuppression with mycophenolate mofetil attenuates hypertension in an experimental model of autoimmune disease," *Journal of the American Heart Association*, vol. 6, no. 3, Article ID e005394, 2017.
- [183] G. E. González, N.-E. Rhaleb, M. A. D'Ambrosio et al., "Deletion of interleukin-6 prevents cardiac inflammation, fibrosis and dysfunctionwithout affecting blood pressure in angiotensin IIhigh salt-induced hypertension," *Journal of Hypertension*, vol. 33, no. 1, pp. 144–152, 2015.
- [184] J. Frieder, D. Kivelevitch, and A. Menter, "Secukinumab: a review of the anti-IL-17A biologic for the treatment of psoriasis," *Therapeutic Advances in Chronic Disease*, vol. 9, no. 1, pp. 5–21, 2018.
- [185] J. Hernández-Sánchez, L. Harlow, C. Church et al., "Clinical trial protocol for TRANSFORM-UK: a therapeutic open-label study of tocilizumab in the treatment of pulmonary arterial hypertension," *Pulmonary Circulation*, vol. 8, no. 1, 2017.
- [186] D. C. Cornelius, J. P. Hogg, J. Scott et al., "Administration of interleukin-17 soluble receptor C supresses TH17 cells, oxidative stress, and hypertension in response to placental ischemia during pregnancy," *Hypertension*, vol. 62, no. 6, pp. 1068–1073, 2013.



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